CASE REPORT

Open Access

Effect of isavuconazole on the concentration of tacrolimus in a patient with genotype *CYP3A5*1/*3*: a case report



Hayato Yokota¹, Yumiko Akamine^{1*}, Harumi Hatakeyama¹, Hideaki Kagaya¹, Sho Sakamoto², Mitsuru Saito³, Masahide Takeda², Kazuhiro Sato², Katsutoshi Nakayama² and Masafumi Kikuchi¹

Abstract

Background Azole antifungals are the standard treatment for pulmonary mycosis, which may develop during long-term immunotherapy for kidney transplant. Isavuconazole (ISCZ) is a cytochrome P450 (CYP) 3 A inhibitor that has a risk of interacting with the immunosuppressive drug tacrolimus (TAC). We report a case of simple pulmonary aspergilloma with renal dysfunction due to increased trough levels of TAC after ISCZ coadministration.

Case presentation A male in his 60s was treated with TAC 3.0 mg/day orally to prevent graft rejection after kidney transplantation. He received a loading dose of ISCZ 600 mg/day orally for two days, followed by a maintenance dose of 200 mg/day for simple pulmonary aspergilloma. The TAC trough concentration increased markedly from 2.4 to 9.9 ng/mL on day 6 after coadministration. The creatinine level increased from 0.70 to 1.08 mg/dL, suggesting renal dysfunction due to TAC. Subsequently, the TAC dosage was reduced, leading to a decreased blood TAC concentration and improved renal function. The patient's genotype was *CYP3A5*1/*3*.

Conclusions In the early stages of ISCZ treatment, the blood TAC concentration is higher, and *CYP3A5* polymorphisms may partially explain the extent of this interaction. We recommend more careful monitoring of TAC and serum creatinine levels for approximately one week after ISCZ administration.

Keywords CYP3A5, Drug-drug interactions, Isavuconazole, Tacrolimus, Therapeutic drug monitoring

*Correspondence:

Yumiko Akamine

yumiko-ai@hos.akita-u.ac.jp

¹Department of Pharmacy, Akita University Hospital, 1-1-1 Hondo, Akita 010-8543, Japan

²Department of Respiratory Medicine, Akita University Graduate School of Medicine, Akita, Japan

³Department of Urology, Akita University Graduate School of Medicine, Akita, Japan

Background

Tacrolimus (TAC), a calcineurin inhibitor, is an immunosuppressive agent used to prevent graft rejection after kidney transplantation [1]. The therapeutic range of blood TAC concentrations is narrow [2, 3], and longterm exposure to high doses can lead to calcineurin inhibitor-induced nephrotoxicity [4]. Therefore, clinicians individually adjust TAC doses based on therapeutic drug monitoring (TDM) to prevent serious adverse events and graft rejection [5].

Patients who undergo kidney transplantation may develop pulmonary mycosis due to prolonged immunosuppressive therapy [6]. In Europe and the United States

© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/jublicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

(US), the recommended first-line therapy for chronic pulmonary aspergillosis (CPA) is azole antifungals, such as voriconazole (VRCZ) and itraconazole (ITCZ) [7, 8]. In Japan, VRCZ and isavuconazole (ISCZ) are each recommended as first-line therapy [9]. ISCZ is the newest triazole antifungal; it was approved by the US Food and Drug Administration and the European Medicines Agency in 2015 to treat invasive aspergillosis and mucormycosis. ISCZ was approved in Japan in 2023 and is also indicated for the treatment of CPA and cryptococcosis. In a randomized, open-label study comparing ISCZ with VRCZ as a control, the overall response rate at the end of treatment for patients with CPA in the ISCZ group and the VRCZ group was 82.7% and 77.8%, respectively [10]. Drug-related adverse events were reported in 61.5% of the ISCZ group and 85.2% of the VRCZ group. Another retrospective study suggested fewer adverse events with ISCZ than with VRCZ [11]. Therefore, this drug is expected to become a new treatment option for fungal infections, including CPA.

Antifungal agents, such as ISCZ, VRCZ, and ITCZ, are known cytochrome P450 (CYP) 3A4/5 inhibitors that induce interactions with TAC, a substrate of CYP3A4/5 [12]. ISCZ has low CYP3A4 inhibitory activity compared with VRCZ and ITCZ [13]. Although ISCZ demonstrated moderate inhibition when combined with TAC, a 2.3fold increase in the area under the TAC blood concentration-time curve occurred with ISCZ coadministration in healthy subjects [14], which may indicate the need for TAC dose adjustment. Previous pharmacokinetic studies have shown that the TAC dose must be reduced to maintain the TAC concentration in the therapeutic range with coadministration of ISCZ [15, 16]. However, marked interpatient variability existed in the degree of drug interaction. The concentration of TAC in the blood exhibits considerable variability among patients [17]. The largely inter-individual variability in the pharmacokinetics of TAC can be explained by a single nucleotide polymorphism (SNP) in CYP3A5 [18]. Zhang Y et al. have shown that in patients who were coadministered VRCZ, the dose-normalized trough concentrations of TAC (C₀/D ratio) were higher in patients with the CYP3A5*3/*3 allele than in those with the CYP3A5*1 allele [19], whereas ISCZ coadministration has not been investigated. Furthermore, little is known about the variability in TAC concentrations during the coadministration of ISCZ in clinical practice in patients with simple pulmonary aspergilloma.

Here, we report a case of a patient with simple pulmonary aspergilloma who showed an increase in blood TAC levels and renal dysfunction with combined administration of TAC and ISCZ. Furthermore, the patient's *CYP3A5* polymorphisms were analyzed.

Case presentation

A man in his 60s was treated with steroids for nephrotic syndrome with membranoproliferative glomerulonephritis 11 years before the current event. The patient underwent kidney transplantation for end-stage renal disease three years prior to the current event. Immunosuppressive treatment consisted of a regimen of TAC, everolimus, mycophenolate mofetil, and prednisolone. Computed tomography performed during treatment revealed an internal cavity in the right upper lobe of the lung containing a mass with a solid nodule within the pulmonary cavity. The patient underwent a bronchoscopy examination after admission. A. fumigatus was detected in the bronchoalveolar lavage fluid, and he was diagnosed with simple pulmonary aspergilloma. After the diagnosis was confirmed, ISCZ was considered as a preoperative treatment. The patient received immunosuppressive maintenance therapy with modified-release formulations of TAC 3 mg/day, mycophenolate mofetil 1000 mg/day, and prednisolone 10 mg/day; everolimus had been discontinued. He also received concomitant bifidobacterium 2 g/day, rabeprazole 10 mg/day, amlodipine 5 mg/ day, sitagliptin 50 mg/day, repaglinide 0.5 mg/day, miglitol 100 mg/day, and lemborexant 5 mg/day. Lemborexant was pre-reduced to 2.5 mg/day because of concerns about interaction with ISCZ. All other drugs were continued at the same dose. Before the administration of ISCZ, the trough blood concentration of TAC was 2.4 ng/mL (target trough level: around 5 ng/mL), and the C_0/D ratio was 0.8 ng/mL/mg, with a creatinine level of 0.70 mg/dL. Oral ISCZ was initiated with a 2-day loading dose of 200 mg three times daily, followed by a daily dose of 200 mg. On day 4 after the start of ISCZ therapy, the trough blood concentration of TAC increased to 6.5 ng/mL (Fig. 1). On day 6, it rose to 9.9 ng/mL (C_0/D ratio: 3.3), which was well above the target concentration. We suspected TAC-induced nephrotoxicity because the creatinine level rose to 0.98 mg/dL. With blood trough levels and laboratory markers indicative of TAC toxicity, the TAC dose was reduced the next day, from 3.0 to 1.5 mg/day. Liver function tests were within the normal range (aspartate transaminase 20 U/L, alanine transaminase 28 U/L, total bilirubin 0.9 mg/dL). The patient was without clinical symptoms and did not require additional treatment. Serum creatinine levels increased to a maximum of 1.08 mg/dL but subsequently decreased after a slight delay following the reduction in TAC dosage. On day 5 after the TAC dose reduction (in Fig. 1, day 11), the TAC blood level decreased to 5.2 ng/mL (C_0/D ratio: 3.5), and the patient was discharged on day 13 (C_0 : 4.4 ng/mL, C_0/D ratio: 2.9 ng/mL). On day 19 after the TAC dose reduction (in Fig. 1, day 25), during an outpatient visit, renal function was observed to have recovered (creatinine: 0.73 mg/dL), and the TAC concentration had

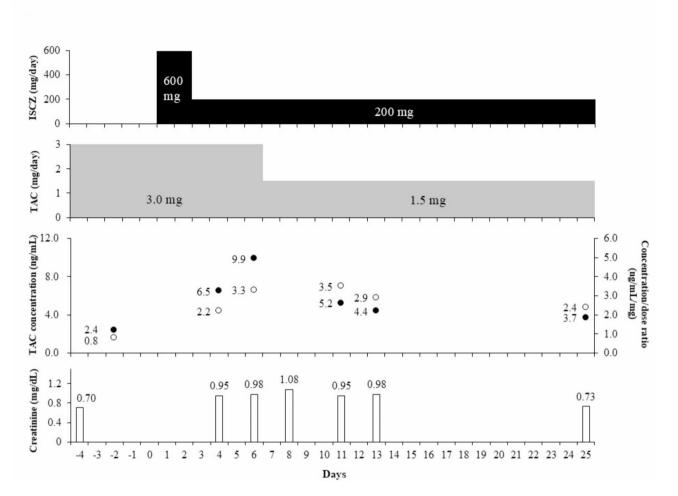


Fig. 1 Clinical course of the patient. The doses of isavuconazole (ISCZ) and tacrolimus (TAC) and blood TAC concentrations (black circles), concentration/ dose ratio (white circles) and laboratory parameters are shown

returned to the rapeutic levels (TAC C_0: 3.7 ng/mL, C_0/D ratio: 2.4).

This case was approved by the Ethics Committee of Akita University School of Medicine (no 1015 and 3207). The patient gave his consent for the publication of this report. According to the manufacturer's instructions, blood TAC concentrations were measured using a chemiluminescence enzyme immunoassay on an Architecti1000 instrument (Abbott Laboratories, Abbott Japan Co. Ltd.). For genotyping of the *CYP3A5* 6986 A > G (*3) SNP, the polymerase chain reaction-restriction fragment length polymorphism method was used [20]. The patient was determined to have a *CYP3A5*1/*3* genotype.

Discussion and conclusions

Here, we report a case of *simple* pulmonary aspergilloma in which drug-drug interactions between ISCZ and TAC led to increased TAC blood levels, followed by the development of renal dysfunction. This case also focused on *CYP3A5* polymorphisms to explore the drug-drug

interaction mechanism. ISCZ is a moderate CYP3A4/3A5 inhibitor [21]. ISCZ has lower CYP3A4 inhibition than VRCZ, ITCZ, and fluconazole in liver microsomes, with an inhibition constant of 0.622 µmol/L [22, 23]. However, the CYP3A4 metabolic pathway for TAC in patients with the CYP3A5*1/*3 genotype, unlike that in patients with the CYP3A5*1/*1 genotype, is expected to be susceptible to CYP3A4/5 inhibition [24, 25]. This suggests that similar results may be observed with the use of ISCZ. Furthermore, in patients with the CYP3A5*3/*3 genotype, the TAC blood concentration increase is higher than that in patients with the CYP3A5*1/*1 and CYP3A5*1/*3 genotypes [19, 24]. The results of this interaction suggest that TAC trough levels are extremely higher with TAC and ISCZ combination treatment in patients with the CYP3A5*3/*3 genotype. Meanwhile, ISCZ is a substrate of CYP3A4 and CYP3A5, with 33.8% and 68.4% residual ISCZ, respectively [21]. Léa Darnaud et al. reported that ISCZ clearance in patients with the CYP3A5*3/*3 genotype is lower than that in patients in the phase 1 and

phase 3 SECURE trials [26, 27]. A further study with more focus on *CYP3A5* polymorphism analysis is necessary to quantify the percentage of TAC dose reductions needed when ISCZ is initiated.

Although the trough concentration of TAC before ISCZ administration tended to be lower than the target concentration, the dose of TAC was not increased, due to concerns about an interaction with ISCZ. A previous study showed that the TAC C₀/D ratio increased 1.44 times on the second day after ISCZ administration [28]. Therefore, in this case, we considered it unlikely that TAC dose adjustments would be necessary within the first two days after ISCZ administration. ISCZ requires a two-day loading dose, and the TAC C_0/D ratio reaches a maximum on the fourth day of coadministration [15]. Thus, we needed to monitor the TAC blood concentration carefully, which was first confirmed on the fourth day of coadministration. In this case, on day 6, the blood TAC concentration and C₀/D ratio increased approximately fourfold from baseline. In a study of patients who had undergone allogeneic hematopoietic stem cell transplantation, the C_0/D ratio of TAC increased within 7 days of ISCZ coadministration, and no significant differences compared to baseline were observed after the second week [29]. In a study of solid organ transplant recipients, in the early stages of ISCZ treatment, a decrease in TAC dosage occurred in 61.3% of patients, but after that, TAC levels remained stable from baseline to one month [30]. Therefore, management of varying blood TAC concentrations is critical for approximately one week after the initiation of coadministration of TAC and ISCZ.

On day four following the initiation of ISCZ treatment, a TAC concentration-dependent increase in creatinine levels was observed, requiring a reduction of the TAC dose by half on day 7. An objective causality assessment based on the Horn Drug Interaction Probability Scale revealed a probable interaction between TAC and ISCZ [31]. High TAC intra-patient variation is associated with a noticeable decrease in renal function and affects CD4+/ CD8+cells, indicators of immune status [32]. Acute kidney injury during hospitalization increases the risk of rehospitalization and increases the risk of overall mortality [33]. Moreover, it has been reported that the TAC maximum trough level was associated with acute kidney injury during admission. Therefore, during TAC administration, we should minimize concentration fluctuations by monitoring TAC trough values to prevent worsening renal function.

According to FDA drug labeling, when VRCZ is used concomitantly with TAC, the TAC dose should be reduced to one-third or lower [34]. In contrast, there is no recommended empirical dose reduction for TAC when coadministered with ISCZ. Fernández-Ruiz M et al. and Monforte A et al. have reported that the daily TAC dose was reduced by 30-50% when ISCZ was coadministered early in treatment [30, 35]. Other studies have suggested that a reduction in the TAC dose of 18% may be recommended [28]. However, the range of TAC dose reductions considered in these reports varies widely, and the reports are not in agreement. Therefore, reducing the TAC dose prophylactically by a defined amount is inappropriate. In the present case, we did not reduce the TAC dose before the start of ISCZ coadministration, but frequent TDM after ISCZ administration may have allowed the patient to continue treatment without discontinuing TAC. Hiratsuka et al. reported that the genotype frequencies of CYP3A5*1/*1, *1/*3, and *3/*3 in Japanese individuals are 7.9, 35.5, and 55.9%, respectively [36]. The allele frequency could support the importance of frequent monitoring of TAC concentrations. Furthermore, CYP3A5 polymorphism analysis may help clinicians improve their understanding of the severity of these interactions. ISCZ is often preferred over other azole antifungals in terms of interactions and is expected to gain more clinical experience in the future.

When initiating administration of a combination of ISCZ and TAC, we recommend that the blood TAC concentration and creatinine levels be carefully monitored for approximately one week. This case shows that the severity of drug interactions may partially be explained by *CYP3A5* polymorphism.

Abbreviations

- C₀/D dose-normalized trough concentration
- CPA chronic pulmonary aspergillosis
- CYP cytochrome P450
- ISCZ isavuconazole
- ITCZ itraconazole
- TAC tacrolimus
- TDM therapeutic drug monitoring
- US United States
- VRCZ voriconazole

Acknowledgements

Not applicable

Author contributions

HY and YA conceptualized and designed the study. HY drafted the manuscript. HY and HH acquired patient data. HK carried out genotyping. SS, MS, KS and MT advised on the interpretation of the therapeutic course in this case and revised the manuscript. KN and MK critically reviewed the manuscript. All authors read and approved the final manuscript.

Funding

This research received no specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Data availability

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

This case was approved by the Ethics Committee of Akita University School of Medicine (no 1015 and 3207). Informed consent was obtained from the patient.

Consent for publication

Informed consent to publish was obtained from the patient presented in this article.

Competing interests

The authors declare no competing interests.

Received: 13 January 2025 / Accepted: 3 March 2025 Published online: 13 March 2025

References

- Scott LJ, McKeage K, Keam SJ, Plosker GL. Tacrolimus: a further update of its use in the management of organ transplantation. Drugs. 2003;63(12):1247– 97. https://doi.org/10.2165/00003495-200363120-00006.
- Staatz CE, Tett SE. Clinical pharmacokinetics of once-daily tacrolimus in solidorgan transplant patients. Clin Pharmacokinet. 2015;54(10):993–1025. https:// doi.org/10.1007/s40262-015-0282-2.
- Staatz CE, Tett SE. Clinical pharmacokinetics and pharmacodynamics of tacrolimus in solid organ transplantation. Clin Pharmacokinet. 2004;43(10):623–53. https://doi.org/10.2165/00003088-200443100-00001.
- Bentata Y, Tacrolimus. 20 Years of use in adult kidney transplantation. What we should know about its nephrotoxicity. Artif Organs. 2020;44(2):140–52. htt ps://doi.org/10.1111/aor.13551.
- Schiff J, Cole E, Cantarovich M. Therapeutic monitoring of calcineurin inhibitors for the nephrologist. Clin J Am Soc Nephrol. 2007;2(2):374–84. https://doi .org/10.2215/cjn.03791106.
- Wilmes D, Coche E, Rodriguez-Villalobos H, Kanaan N. Fungal pneumonia in kidney transplant recipients. Respir Med. 2021;185:106492. https://doi.org/10. 1016/j.rmed.2021.106492.
- Patterson TF, Thompson GR 3rd, Denning DW, Fishman JA, Hadley S, Herbrecht R, et al. Practice guidelines for the diagnosis and management of aspergillosis: 2016 update by the infectious diseases society of America. Clin Infect Dis. 2016;63(4):e1–60. https://doi.org/10.1093/cid/ciw326.
- Denning DW, Cadranel J, Beigelman-Aubry C, Ader F, Chakrabarti A, Blot S et al. Chronic pulmonary aspergillosis: rationale and clinical guidelines for diagnosis and management. Eur Respir J. 2016;47(1):45–68. intro. https://doi. org/10.1183/13993003.00583-2015
- JAID/JSC Guide/Guidelines to Clinical Management of Infectious Disease Preparing Committee. The JAID/JSC guide to clinical management of infectious diseases 2023. Tokyo: Japanese Association for Infectious Disease/Japanese Society of Chemotherapy; 2023.
- Kohno S, Izumikawa K, Takazono T, Miyazaki T, Yoshida M, Kamei K, et al. Efficacy and safety of isavuconazole against deep-seated mycoses: A phase 3, randomized, open-label study in Japan. J Infect Chemother. 2023;29(2):163– 70. https://doi.org/10.1016/j.jiac.2022.10.010.
- Bongomin F, Maguire N, Moore CB, Felton T, Rautemaa-Richardson R. Isavuconazole and voriconazole for the treatment of chronic pulmonary aspergillosis: A retrospective comparison of rates of adverse events. Mycoses. 2019;62(3):217–22. https://doi.org/10.1111/myc.12885.
- Groll AH, Townsend R, Desai A, Azie N, Jones M, Engelhardt M, et al. Drugdrug interactions between Triazole antifungal agents used to treat invasive aspergillosis and immunosuppressants metabolized by cytochrome P450 3A4. Transpl Infect Dis. 2017;19(5). https://doi.org/10.1111/tid.12751.
- Chau MM, Daveson K, Alffenaar JC, Gwee A, Ho SA, Marriott DJE, et al. Consensus guidelines for optimising antifungal drug delivery and monitoring to avoid toxicity and improve outcomes in patients with haematological malignancy and Haemopoietic stem cell transplant recipients, 2021. Intern Med J. 2021;51(Suppl 7):37–66. https://doi.org/10.1111/imj.15587.
- Groll AH, Desai A, Han D, Howieson C, Kato K, Akhtar S, et al. Pharmacokinetic assessment of drug-drug interactions of isavuconazole with the immunosuppressants cyclosporine, mycophenolic acid, prednisolone, sirolimus, and tacrolimus in healthy adults. Clin Pharmacol Drug Dev. 2017;6(1):76–85. https: //doi.org/10.1002/cpdd.284.

- Rivosecchi RM, Clancy CJ, Shields RK, Ensor CR, Shullo MA, Falcione BA, et al. Effects of isavuconazole on the plasma concentrations of tacrolimus among solid-organ transplant patients. Antimicrob Agents Chemother. 2017;61(9):e00970–17. https://doi.org/10.1128/aac.00970-17.
- Trifilio S, Rubin H, Monacelli A, Mehta J. Tacrolimus dose modification in patients receiving concomitant isavuconazole after hematopoietic stem cell transplantation. J Oncol Pharm Pract. 2021;27(4):857–62. https://doi.org/10.1 177/1078155220940416.
- Venkataramanan R, Swaminathan A, Prasad T, Jain A, Zuckerman S, Warty V, et al. Clinical pharmacokinetics of tacrolimus. Clin Pharmacokinet. 1995;29(6):404–30. https://doi.org/10.2165/00003088-199529060-00003.
- Coto E, Tavira B, Suárez-Álvarez B, López-Larrea C, Díaz-Corte C, Ortega F et al. Pharmacogenetics of tacrolimus: ready for clinical translation? Kidney Int Suppl (2011). 2011;1(2):58–62. https://doi.org/10.1038/kisup.2011.14
- Zhang Y, Du Y, Ren S, Li Y, Zhang X, Cao X, et al. CYP3A5 genotype-dependent drug-drug interaction between tacrolimus and voriconazole in Chinese kidney transplant patients. Ann Pharmacother. 2024;58(6):605–13. https://doi. org/10.1177/10600280231197399.
- Fukuen S, Fukuda T, Maune H, Ikenaga Y, Yamamoto I, Inaba T, et al. Novel detection assay by PCR-RFLP and frequency of the CYP3A5 SNPs, CYP3A5*3 and *6, in a Japanese population. Pharmacogenetics. 2002;12(4):331–4. https: //doi.org/10.1097/00008571-200206000-00009.
- 21. Asahi Kasei Pharma Corporation. The drug interview form of Isavuconazonium Sulfate (Cresemba[®]) [Internet]. [cited 2024 Sep 14]. Available from: https ://akp-pharma-digital.com/products/list/6498?btd_ref
- Yamazaki H, Nakamoto M, Shimizu M, Murayama N, Niwa T. Potential impact of cytochrome P450 3A5 in human liver on drug interactions with Triazoles. Br J Clin Pharmacol. 2010;69(6):593–7. https://doi.org/10.1111/j.1365-2125.20 10.03656.x.
- Asahi Kasei Pharma Corporation. Cresemba Capsules 100 mg /Cresemba for i.v. infusion 200 mg. CTD 2.7.2 Summary of Clinical Pharmacology Studies, 2.7.2.2 Summary of Results of Individual Studies [Internet]. [cited 2024 Sep 14]. Available from: https://www.pmda.go.jp/drugs/2023/P20230118002/ind ex.html
- Nara M, Takahashi N, Miura M, Niioka T, Kagaya H, Fujishima N, et al. Effect of Itraconazole on the concentrations of tacrolimus and cyclosporine in the blood of patients receiving allogeneic hematopoietic stem cell transplants. Eur J Clin Pharmacol. 2013;69(6):1321–9. https://doi.org/10.1007/s00228-01 3-1471-2.
- Chandel N, Aggarwal PK, Minz M, Sakhuja V, Kohli KK, Jha V. CYP3A5*1/*3 genotype influences the blood concentration of tacrolimus in response to metabolic Inhibition by ketoconazole. Pharmacogenet Genomics. 2009;19(6):458–63. https://doi.org/10.1097/FPC.0b013e32832bd085.
- Darnaud L, Lamoureux F, Godet C, Pontier S, Debard A, Venisse N, et al. Isavuconazole kinetic exploration for clinical practice. Drugs R D. 2018;18(4):317– 21. https://doi.org/10.1007/s40268-018-0251-y.
- Desai A, Kovanda L, Kowalski D, Lu Q, Townsend R, Bonate PL. Population pharmacokinetics of isavuconazole from phase 1 and phase 3 (SECURE) trials in adults and target attainment in patients with invasive infections due to Aspergillus and other filamentous fungi. Antimicrob Agents Chemother. 2016;60(9):5483–91. https://doi.org/10.1128/aac.02819-15.
- Fructuoso-González L, Najera-Perez MD, Manresa-Ramón N, Torrano-Belmonte P, Caracena-López S, Pacheco-López P. Isavuconazole-tacrolimus drug-drug interactions in HSCT patients. J Antimicrob Chemother. 2023;78(10):2559–62. https://doi.org/10.1093/jac/dkad271.
- Kieu V, Jhangiani K, Dadwal S, Nakamura R, Pon D. Effect of isavuconazole on tacrolimus and sirolimus serum concentrations in allogeneic hematopoietic stem cell transplant patients: A drug-drug interaction study. Transpl Infect Dis. 2019;21(1):e13007. https://doi.org/10.1111/tid.13007.
- Fernández-Ruiz M, Bodro M, Gutiérrez Martín I, Rodriguez-Álvarez R, Ruiz-Ruigómez M, Sabé N, et al. Isavuconazole for the treatment of invasive mold disease in solid organ transplant recipients: a multicenter study on efficacy and safety in real-life clinical practice. Transplantation. 2023;107(3):762–73. htt ps://doi.org/10.1097/tp.00000000004312.
- Horn JR, Hansten PD, Chan LN. Proposal for a new tool to evaluate drug interaction cases. Ann Pharmacother. 2007;41(4):674–80. https://doi.org/10.1 345/aph.1H423.
- Wang X, Liu Z, Chen J, Chai Y, Shao X, Xie W, et al. Impact of intra-patient variability of tacrolimus on allograft function and CD4 + /CD8 + ratio in kidney transplant recipients: a retrospective single-center study. Int J Clin Pharm. 2024;46(4):918–25. https://doi.org/10.1007/s11096-024-01726-w.

- Hod T, Oberman B, Scott N, Levy L, Shlomai G, Beckerman P, et al. Predictors and adverse outcomes of acute kidney injury in hospitalized renal transplant recipients. Transpl Int. 2023;36:11141. https://doi.org/10.3389/ti.2023.11141.
- Food and Drug Administration. VFEND® (Voriconazole) Tablets, for Oral Use and for Intravenous Use. [Internet]. 2021 [cited 2024 Sep 1]. Accessed September 1. 2024. Available from: https://www.accessdata.fda.gov/scripts/cder/ daf/index.cfm?event=overview.process%26;AppINo=021266
- Monforte A, Los-Arcos I, Martín-Gómez MT, Campany-Herrero D, Sacanell J, Berastegui C, et al. Safety and effectiveness of isavuconazole treatment for fungal infections in solid organ transplant recipients (ISASOT Study). Microbiol Spectr. 2022;10(1):e0178421. https://doi.org/10.1128/spectrum.01784-21.
- Hiratsuka M, Takekuma Y, Endo N, Narahara K, Hamdy SI, Kishikawa Y, et al. Allele and genotype frequencies of CYP2B6 and CYP3A5 in the Japanese population. Eur J Clin Pharmacol. 2002;58(6):417–21. https://doi.org/10.1007/ s00228-002-0499-5.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.